

UC Irvine

UC Irvine Previously Published Works

Title

Cost and Scalability Analysis of Porcine Islet Isolation for Islet Transplantation: Comparison of Juvenile, Neonatal and Adult Pigs.

Permalink

<https://escholarship.org/uc/item/22s14102>

Journal

Cell transplantation, 28(7)

ISSN

0963-6897

Authors

Vanderschelden, Rachel
Sathialingam, Mayilone
Alexander, Michael
[et al.](#)

Publication Date

2019-07-01

DOI


10.1177/0963689719847460

Copyright Information

This work is made available under the terms of a Creative Commons Attribution License, available at <https://creativecommons.org/licenses/by/4.0/>

Peer reviewed

Cost and Scalability Analysis of Porcine Islet Isolation for Islet Transplantation: Comparison of Juvenile, Neonatal and Adult Pigs

Cell Transplantation
2019, Vol. 28(7) 967–972
© The Author(s) 2019
Article reuse guidelines:
sagepub.com/journals-permissions
DOI: 10.1177/0963689719847460
journals.sagepub.com/home/cll


Rachel Vanderschelden^{1,2}, Mayilone Sathialingam²,
Michael Alexander^{1,2,3}, and Jonathan R. T. Lakey^{1,2,3} 

Abstract

The limited availability of human islets has led to the examination of porcine islets as a source of clinically suitable tissue for transplantation in patients with diabetes mellitus. Islets from porcine donors are commonly used in both *in vitro* and *in vivo* experiments studying diabetes mellitus. However, there are significant differences in quality and quantity of islet yield depending on donor pig age, as well as substantial differences in the costs of pancreas procurement in adult versus neonatal and juvenile pigs. In this study, we compared the total cost per islet of juvenile pig pancreata with that of neonatal and adult pigs. Although adult porcine pancreata yield, on average, more than five times the amount of islets than do juvenile and neonatal pancreata, we found that the high price of adult pigs led to the cost per islet being more than twice that of juvenile and neonatal islets (US \$0.09 vs \$0.04 and \$0.02, respectively). In addition, neonatal and juvenile islets are advantageous in their scalability and retention of viability after culture. Our findings indicate that isolating neonatal and juvenile porcine islets is more cost-effective and scalable than isolating adult porcine islets.

Keywords

type 1 diabetes mellitus, scalability, neonatal porcine islets, juvenile porcine islets

Introduction

Development of the Edmonton Protocol in 2001 and demonstration of the viability of islet cell transplantation as treatment for type 1 diabetes mellitus has led to increased interest in optimization of human islet transplantation for treatment of this disease^{1,2}. However, limited availability of viable human islets and costs of human islet isolation and characterization creates an obstacle for widespread use of islet transplantation for diabetes treatment and has prompted the examination of porcine islets as a clinically suitable substitute¹.

At present, adult pigs are considered superior in many respects to young (neonatal and juvenile) pigs as a source of islets for xenotransplantation. Young porcine pancreata contain islet-like cell clusters which contain sparse insulin-producing cells in comparison to mature islets³. Thus, separation of insulin-producing cells from exocrine cells is more difficult in young pigs^{4–6}. Previous studies have demonstrated that adult porcine islets have greater insulin secretion rates than neonatal and juvenile porcine islets⁷.

However, there are potential benefits to utilizing young pig islets, including reduced immunogenicity, increased cell proliferation ability post-transplantation, and increased *in vitro* stability versus adult islets^{8–10}. Furthermore, neonatal islets have been shown to be effective in restoring normoglycemia after transplantation into diabetic mice¹¹. Isolation of adult porcine islets is also labor-intensive and

¹ Sue and Bill Gross Stem Cell Research Center, University of California, Irvine, USA

² Department of Surgery, University of California, Irvine, USA

³ Department of Biomedical Engineering, University of California, Irvine, USA

Submitted: November 5, 2018. Revised: April 5, 2019. Accepted: April 8, 2019.

Corresponding Author:

Jonathan R. T. Lakey, Department of Surgery and Biomedical Engineering, University of California Irvine, 333 City Boulevard West, Suite 1600, Orange, CA 92868, USA.

Email: jlakey@uci.edu



Creative Commons Non Commercial CC BY-NC: This article is distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 License (<http://www.creativecommons.org/licenses/by-nc/4.0/>) which permits non-commercial use, reproduction and distribution of the work without further permission provided the original work is attributed as specified on the SAGE and Open Access pages (<https://us.sagepub.com/en-us/nam/open-access-at-sage>).

expensive, especially when considering the prolonged husbandry costs required to produce adult pigs. Therefore, utilization of young pigs may potentially mitigate some of these costs^{2,12}.

No previous study has compared the cost-effectiveness of neonatal, juvenile, and adult porcine islet isolations. Smith et al. had previously compared the oxygen demand, membrane integrity, and cell function and proliferation between neonatal, juvenile, and adult porcine islets⁷. However, the pricing to carry out each group of experiments was not considered. Therefore, there is a need to understand the cost-effectiveness and scalability of each age group of pigs to facilitate understanding of their roles in islet isolation and xenotransplantation.

To accomplish this objective, we developed a scalable protocol for porcine islet isolation using 18- to 24-day-old juvenile pigs that has shown promise in *in vivo* transplant studies in rodent models of diabetes. The aim of this study was to determine the actual costs to produce islets from our pre-weaned 18- to 24-day-old juvenile pigs and compare these with the costs of producing islets in neonatal (3 days old) and adult pigs, as reported in the current literature. We reviewed several studies investigating islet yield in adult and neonatal pigs and determined the price per viable islet equivalent (IEQ) of each group of pigs in the context of their respective advantages and disadvantages.

Materials and Methods

Islet Isolation of Juvenile Pigs

Islets were isolated from 18- to 24-day-old male Yorkshire swine ($n = 12$; S&S Farms, Ramona, CA, USA) using procedures we described previously¹³. Briefly, the pancreas was harvested using rapid surgical procurement (<5 min) and placed in Organ Preservation Solution (Corning Cellgro, Manassas, VA, USA). Cold ischemia time was limited to <30 min. The pancreas was then washed in cold (4°C) Hanks balanced salt solution supplemented with HEPES (Corning Cellgro, Manassas, VA, USA), and trimmed of surrounding adipose and lymphatic tissue in a sterile biosafety hood. The pancreatic tissue was then minced into 2–3 mm² pieces and digested at 37°C using Sigma Type V Collagenase (2.5 mg/mL in Hanks balanced salt solution; Sigma Aldrich, St. Louis, MO, USA). The islet tissue clusters (50–500 µm) isolated using this method were allowed to mature into islet cell clusters during *in vitro* culture at 37°C, 5% CO₂ first in Recovery Maturation Media (University of California Irvine, Irvine, CA, USA) supplemented with 215 mM aprotinin (Sigma-Aldrich), 0.5 mM Pefabloc (Sigma-Aldrich), 417 mM dornase alfa (Genentech, San Francisco, CA, USA), and 10% porcine serum (Omega Scientific, Tarzana, CA, USA), and 48 h later, in a novel maturation media (University of California Irvine, Irvine, CA, USA) supplemented with 10% porcine serum (Omega Scientific, Tarzana, CA, USA) as outlined previously¹³.

Yield, Viability and Function Determination

Juvenile pigs. Islet yield was calculated from the previous 12 juvenile pig islet isolations. At the end of the 7-day culture period, islet quality control assessment was performed. Islet count and IEQ were determined by staining a 100-µL aliquot with 1 mL dithizone (MP Biomedicals, Santa Ana, CA, USA), and observing islets at 25x on a standard stereomicroscope (Max Erb, Santa Ynez, CA, USA) containing a 10x eye piece graticule. Islet viability was analyzed using fluorescein diacetate (FDA) and propidium iodide (PI) (Invitrogen, Carlsbad, CA, USA), imaged using fluorescence microscopy (Nikon LSM510; Thornwood, NY, USA), and quantified with a Microplate reader (Tecan Infinite F200, Magellan V7; Tecan, Männedorf, Switzerland).

Islet function was determined using glucose-stimulated insulin release; 100–150 porcine islets were incubated for 1 h, in corresponding order, in low glucose (2.8 mM), high glucose (28 mM), high glucose plus 3-isobutyl-1-methylxanthine (28 mM, 50 µM), and then low glucose (D-(+)-glucose; Sigma Aldrich, St. Louis, MO, USA); stimulation index (SI) was calculated as the ratio of insulin secreted in high glucose over the amount of insulin secreted in low glucose. Insulin levels were measured using a standard porcine insulin enzyme-linked immunosorbent assay (Porcine Insulin ELISA; Mercodia, Winston Salem, NC, USA), and absorbance was measured using a Microplate reader (InfiniteF200 and Magellan V7; Tecan, Männedorf, Switzerland).

Adult pigs. Average yield and viability of adult islets were estimated from the literature. Four studies composed of nine subgroups and a total of 131 adult pigs were analyzed (Table 1)^{4–5,14–15}. One study reported viability for each of its three subgroups, and average viability was calculated from this study. SI, as measured by glucose-stimulated insulin release, was reported in two studies.

Neonatal pigs. Average yield and viability of neonatal islets were estimated from the literature. One study comprised of three subgroups and 136 pigs was analyzed¹⁶. Average SI, as measured by glucose-stimulated insulin release, was reported for each subgroup (Table 2). Viability data was not available and thus not considered for neonatal pigs in this paper.

Cost Determination

For neonatal, preweaned juvenile and adult pigs, islet isolation cost was divided into three categories. Animal source, husbandry, and veterinary costs were grouped together as “Husbandry”, surgical procurement and organ preservation including operating room costs were grouped as “Surgical”, and isolation and tissue culture costs were grouped as “Isolation”. All categories included applicable materials as well as labor cost. Isolation and surgical costs were calculated using values provided by the University of California,

Table 1. Average Yield, Viability, Stimulation Index (SI), and Cost of Adult Porcine Islets.

Study	Subgroup	n	Average Yield ($\times 10^3$ IEQ) ^a	Viability (%)	SI ^a	Cost per IEQ (\$)
Yonekawa	New Method	5	880 \pm 223	82	1.2 \pm 0.2	0.034
	Ricordi Method	5	317 \pm 86	94	1.6 \pm 0.3	0.095
	Open Pan Method	5	30.6 \pm 11.5	84	1.2 \pm 0.2	0.98
Bottino	Ricordi Method	14	597 \pm 60	NR	2.2 \pm 0.5	0.050
Anazawa	Group 1	8	335 \pm 114	NR	NR	0.90
	Group 2	12	366 \pm 101	NR	NR	0.082
	Group 3	14	491 \pm 188	NR	NR	0.061
Kim	High Yield	34	305 \pm 18.2	NR	NR	0.098
	Low Yield	34	143 \pm 12.5	NR	NR	0.210
Overall		131	217 \pm 327	86.6	—	0.090

^aMean values \pm SD. NR: Not reported**Table 2.** Average Yield and Stimulation Index of Neonatal Porcine Islets.

Study	Subgroup	n	Average Yield ($\times 10^3$ IEQ) ^a	Viability (%)	SI ^a
Korbutt	Group A	64	35.1 \pm 0.135	NR	4.3 \pm 0.2
	Group B	24	38.6 \pm 0.254	NR	4.4 \pm 0.2
	Group C	48	41.8 \pm 0.226	NR	5.5 \pm 0.1
Overall		136	38.1 \pm 3.0	—	4.7 \pm 0.58

^aMean values \pm SD. NR: Not reported**Table 3.** Neonatal, Juvenile, and Adult Islet Isolation—Complete Costs Per Pig.

Category	Neonatal	Juvenile	Adult
Husbandry	\$96.01	\$576.04	\$9,958.78
Surgical	\$30.50	\$30.50	\$228.77
Isolation	\$642.76	\$642.76	\$16,946.40
Sum	\$769.26	\$1,249.29	\$27,133.94

Irvine (CA, USA). An institutional tax of 29.8% was applied across all categories (Table 3).

Data Analysis

The primary statistical endpoints of this study were islet yield, viability, and SI of each population. Pooled means and standard deviations (SDs) were appropriately calculated when the given population was homogeneous. For instance, data for juvenile pigs was obtained from a single protocol and population, and data for neonatal pigs was obtained from a single study. Thus, these populations were each treated as homogeneous. For juvenile pigs, cost per islet was determined using this pooled mean and the calculated cost of juvenile islet procurement. Similarly, viability as well as mean and SD of SI was determined from pooled data for the 12 trials of this study.

For neonatal pigs, a pooled mean and SD was calculated under the assumption that subgroups from the single analyzed study originated from a homogeneous population. Cost per islet was determined using this pooled mean and the calculated cost of neonatal islet procurement. Similarly,

mean and SD of SI was determined from pooled data for the three subgroups of this study. Viability data was not available.

Data regarding islet yield in adult porcine pancreata was extracted from previous studies^{4–5,14–15}. However, because studies were conducted under differing conditions and samples were not homogenous, pooled means and SDs are limited in their usefulness of analyzing this data¹⁷. Instead, each study was evaluated individually and the cost per islet was determined based on the number of islets procured in each study and the calculated cost of adult porcine pancreata (Table 1). For illustrative purposes, and in order to broadly compare average islet yields in adult versus juvenile and neonatal pigs, a simple weighted average of islet yield was calculated to account for differences in sample size among the included studies. Viability was calculated using simple weighted averages and pooled SDs, as these values arose from a single study and thus the population was deemed to be homogenous. SI was reported in two studies, and thus pooling was not appropriate. Indices are reported in Table 1.

Results

Yield, Viability and Function of Islets in Juvenile, Adult, and Neonatal Pigs

Islet yield after 7 days of *in vitro* culture was 30.4 \pm 1.20 $\times 10^3$ IEQ per juvenile pig, with average viability of 87.5%. Glucose-stimulated insulin response gave an average value of 1.77 \pm 0.49.

Values from nine protocols and 131 animals were used to determine the range of islet yields from adult porcine

Table 4. Average Yield, Price, Viability and Stimulation Index by Age of Donor.

	Average Yield/Pancreas ($\times 10^3$ IEQ) ^a	Price/IEQ (\$)	Average Viability (%)	Average SI
Neonatal	38.1 \pm 3.0	0.02	—	4.7 \pm 0.58
Juvenile	30.4 \pm 1.2	0.04	87.5	1.77 \pm 0.49
Adult	333 \pm 129	0.09	86.7	1.75 \pm 0.60

^aMean values \pm SD.

pancreata. Islet yield averages ranged from 30×10^3 to 880×10^3 IEQ per pig with viabilities ranging from 82% to 94%, and SIs ranging from 1.2 to 2.2 (Table 1). For illustrative purposes, average islet yield was determined to be $333 \pm 129 \times 10^3$ IEQ per adult pig. Average reported viability was 86.7% (Table 1).

Values from one protocol (three subgroups) and 136 animals were used to determine the average islet yield from neonatal porcine pancreata. Average islet yield was determined to be $38.1 \pm 3.0 \times 10^3$ IEQ per adult pig with an average reported viability of 87.5%. Average SI was calculated to be 4.7 ± 0.58 (Table 2).

Cost Comparison of Juvenile, Adult, and Neonatal Pig Islet Procurement

There were no differences in isolation or surgical costs for neonatal or juvenile pigs at our institution. Costs of adult pig husbandry were US \$9958 (costs for raising pigs raised to 2 years of age) compared with approximately \$96 for neonatal and \$576 for 18- to 24-day-old pre-weaned juvenile pigs (Table 3). Surgical costs were also moderately less expensive for neonatal and juvenile pigs (\$30.50 per animal) compared with adult pigs (\$228.77 per animal). Notably, up to 12–15 juvenile pig pancreases can be recovered in the same time it would require to complete 1 adult pig pancreas procurement (1.5–2 h). Isolation costs for neonatal and juvenile pigs were calculated at \$642.76 per isolation, including 7 days *in vitro* tissue culture, compared with approximately \$16,946 for adult pig isolation including gradient purification and *in vitro* tissue culture. Therefore, the total cost of neonatal islet isolation was calculated as \$769 per pancreas, juvenile islet isolation as \$1249 per pancreas, and adult islet isolation as \$27,133 per pancreas. The price per IEQ was therefore determined to be \$0.02 for neonatal pigs and \$0.04 for juvenile pigs. Price per IEQ for adult pigs ranged from \$0.03 to nearly \$1.00. An average price per IEQ of \$0.09 was estimated for adult pigs (Table 4, Supplemental Table 5).

Discussion and Conclusion

Study Limitations

There were a few notable limitations to this study owing to its retrospective nature. Because our laboratory does not

isolate islets from adult or neonatal porcine pancreata, data regarding these subgroups had to be estimated from the literature. Although we were able to find data from multiple studies examining adult porcine islet yields, there were few studies that reported this data in neonatal piglets, and thus sample size was limited in this group. Furthermore, although pooling multiple studies on adult islet isolation allowed for a more complete analysis of islet yields in this population, it also limits the efficacy of a pooled mean and SD in this group, as the populations of these studies cannot be assumed to be identical. Since the studies all sampled adult pigs, and we were not concerned with specific method of isolation, we were able to generate a pooled mean for limited illustrative purposes. However, the cost per IEQ of each individual study is a more exact measure of this value (Table 1).

Cost Effectiveness and Scalability of Juvenile, Neonatal, and Adult Porcine Islets

In this study, we evaluated the cost-efficiency and scalability of neonatal, juvenile, and adult porcine islets, accounting for husbandry, surgery, and isolation costs for each group. Our findings indicate that neonatal (\$0.02/IEQ) and juvenile (\$0.04/IEQ) porcine islets are more cost-effective than adult porcine islets (\$0.09/IEQ, range \$0.03 to \$0.98). Looking solely at cost, juvenile porcine islets cost less than half of the price of adult islets while having nearly identical viability and SI measurements. Neonatal porcine islets had more than double the SI of juvenile and adult islets at only a fraction of the cost. However, there was limited literature that specified neonatal porcine islet yield, and no study reported viability in the data we gathered. Nevertheless, previous studies have confirmed higher stimulation indices in neonatal porcine islets versus adult porcine islets, attributing their findings to the lower basal secretion of neonatal islets and the minimal damage sustained during the less aggressive isolation procedure. The 7-day culture of neonatal and juvenile islets also may allow for better cell recovery and function, since the adult islets are not cultured prior to secretion assays^{18–19}. Moreover, while islet yields in young pigs have been shown to increase throughout a culture period, likely due to further differentiation of duct cells into islet cells, studies have shown a 30–50% decrease in yield after culture of adult islets^{14,19}. Furthermore, studies suggest that neonatal islets continue to secrete insulin beyond the time that the secretory function of adult islets begins to decline²⁰.

The reproducibility and scalability of juvenile and neonatal porcine islet isolations is superior to that of adult porcine islets. Adult pig isolations had highly variable total islet yields (Table 1), whereas neonatal and juvenile pig isolations had relatively consistent total islet yields, thereby indicating their increased reproducibility. Part of the variability of the adult porcine islet yields can be attributed to differing isolation conditions; the success of such adult isolations have been shown to depend on morphological screening of the pancreas, warm ischemia time, and endotoxin content,

among other factors²¹. In terms of scalability, the relative simplicity of the juvenile and neonatal porcine islet isolation procedure maximizes islet yield per hour of labor, since the procedure essentially consists of removing the pancreas and chopping it into small fragments, followed by enzymatic digestion and subsequent cell culture. However, adult porcine islet isolation is much more complicated. In short, one must first inject collagenase selectively through the pancreatic duct and later break down fibrous tissue by placing the pancreas within a Ricordi chamber and continue to vigorously shake it while also being careful not to damage the islets²². This aggressive isolation procedure can limit islet yield, function, and viability through islet destruction. Recent studies have begun using an Oxford chamber—a digestion and filtration chamber similar to the Ricordi chamber—because it results in reduced tissue destruction and greater islet yields²³. Nevertheless, the use of both chambers requires more training and a more optimized protocol due to the many variables that can alter isolation success, like collagenase injection pressure, digestion time, and chamber temperature^{24,25}.

As a result of the relative ease of the juvenile and neonatal pig isolations, under our juvenile pancreatic isolation procedure, as many as 24 pancreases have been procured over a 4-h period, illustrating the improved scalability of the procedure when compared with adult pig pancreas isolations. Even at this heightened pace, juvenile islet yields were similar to results obtained with standard processing of two to four pancreases over the same period of time. Juvenile islet isolation also requires less extensive training and decreased labor hours, taking 20 h to isolate from four juvenile pigs, but 58 h to isolate from 1 adult pig. These islet isolation times were determined from our isolation data of juvenile pigs and data provided by the University of California, Irvine. This disparity was also accounted for in our cost calculation per IEQ for each porcine islet group.

Our findings indicate that isolating neonatal and juvenile porcine islets is more cost-effective and scalable than isolating adult porcine islets. Neonatal and juvenile porcine islets demonstrated similar or superior viability and SI values when compared with adult porcine islets. Furthermore, the adult porcine islet isolation procedure is more difficult and labor-intensive to successfully conduct and produce consistent results, making the juvenile and neonatal porcine islet isolations more scalable for future experimentation.

Ethics Approval

All procedures in this study were conducted in accordance with University of California Irvine Institute of Animal Care and Use Committee, approved protocol #AUP-17-129.

Statement of Human and Animal Rights

All procedures were performed according to protocols approved by the University of California Irvine Institute of Animal Care and Use Committee.

Statement of Informed Consent

Informed consent is not applicable because there are no human subjects in this study.


Declaration of Conflicting Interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Funding

The author(s) received no financial support for the research, authorship, and/or publication of this article.

ORCID iD

Jonathan R. T. Lakey  <https://orcid.org/0000-0001-8553-4287>

Supplemental Material

Supplemental material for this article is available online.

References

1. Ellis CE, Korbitt GS. Justifying clinical trials for porcine islet xenotransplantation. *Xenotransplantation*. 2015;22(5):336–344.
2. Park CG, Bottino R, Hawthorne WJ. Current status of islet xenotransplantation. *Int J Surg*. 2015;23(Pt B):261–266.
3. Korsgren O, Christofferson R, Jansson L. Angiogenesis and angioarchitecture of transplanted fetal porcine islet-like cell clusters. *Transplantation*. 1999;68(11):1761–1766.
4. Kim HI, Lee SY, Jin SM, Kim KS, Yu JE, Yeom SC, Yoon TW, Kim JH, Ha J, Park CG, Kim SJ. Parameters for successful pig islet isolation as determined using 68 specific-pathogen-free miniature pigs. *Xenotransplantation*. 2009;16(1):11–18.
5. Bottino R, Balamurugan AN, Smetanka C, Bertera S, He J, Rood PPM, Cooper DKC, Trucco M. Isolation outcome and functional characteristics of young and adult pig pancreatic islets for transplantation studies. *Xenotransplantation*. 2007;14(1):74–82.
6. Alumets J, Håkanson R, Sundler F. Ontogeny of endocrine cells in porcine gut and pancreas. An immunocytochemical study. *Gastroenterology*. 1983;85(6):1359–1372.
7. Smith KE, Purvis WG, Davis MA, Min CG, Cooksey AM, Weber CS, Jandova J, Price ND, Molano DS, Stanton JB, Kelly AC, Steyn LV, Lynch RM, Limesand SW, Alexander M, Lakey JRT, Seeberger K, Korbitt GS, Mueller KR, Hering BJ, McCarthy FM, Papas KK. In vitro characterization of neonatal, juvenile, and adult porcine islet oxygen demand, β -cell function, and transcriptomes. *Xenotransplantation*. 2018;25(6):e12432.
8. Bloch K, Assa S, Lazard D, Abramov N, Shalitin S, Weintrob N, Josefsberg Z, Rapoport M, Vardi P. Neonatal pig islets induce a lower T-cell response than adult pig islets in IDDM patients. *Transplantation*. 1999;67(5):748–752.
9. Trivedi N, Hollister-Lock J, Lopez-Avalos MD, O’Neil JJ, Keegan M, Bonner-Weir S, Weir GC. Increase in beta-cell mass in transplanted porcine neonatal pancreatic cell clusters is due to proliferation of beta-cells and differentiation of duct cells. *Endocrinology*. 2001;142(5):2115–2122.

10. Yoon KH, Quicquel RR, Tatarkiewicz K, Ulrich TR, Hollister-Lock J, Trivedi N, Bonner-Weir S, Weir GC. Differentiation and expansion of beta cell mass in porcine neonatal pancreatic cell clusters transplanted into nude mice. *Cell Transplant*. 1999;8(6):673–689.
11. Weir GC, Quicquel RR, Yoon KH, Tatarkiewicz K, Ulrich TR, Hollister-Lock J, Bonner-Weir S. Porcine neonatal pancreatic cell clusters (NPCCs): a potential source of tissue for islet transplantation. *Ann Transplant*. 1997;2(3):63–68.
12. Nagaraju S, Bottino R, Wijkstrom M, Trucco M, Cooper DKC. Islet xenotransplantation: what is the optimal age of the islet-source pig? *Xenotransplantation*. 2015;22(1):7–19.
13. Lamb M, Laugenour K, Liang O, Alexander M, Foster CE, Lakey JRT. In vitro maturation of viable islets from partially digested young pig pancreas. *Cell Transplant*. 2014;23(3):263–272.
14. Anazawa T, Balamurugan AN, Papas KK, Avgoustiniatos ES, Ferrer J, Matsumoto S, Sutherland DER, Hering BJ. Improved method of porcine pancreas procurement with arterial flush and ductal injection enhances islet isolation outcome. *Transplant Proc*. 2010;42(6):2032–2035.
15. Yonekawa Y, Matsumoto S, Okitsu T, Arata T, Iwanaga Y, Noguchi H, Nagata H, O'Neil JJ, Tanaka K. Effective islet isolation method with extremely high islet yields from adult pigs. *Cell Transplant*. 2005;14(10):757–762.
16. Ellis C, Lyon JG, Korbitt GS. Optimization and scale-up isolation and culture of neonatal porcine islets: potential for clinical application. *Cell Transplant*. 2016;25(3):539–547.
17. Bangdiwala SI, Bhargava A, O'Connor DP, Robinson TN, Michie S, Murray DM, Stevens J, Belle SH, Templin TN, Pratt CA. Statistical methodologies to pool across multiple intervention studies. *Transl Behav Med*. 2016;6(2):228–235.
18. Cooper DKC, Matsumoto S, Abalovich A, Itoh T, Mourad NI, Gianello PR, Wolf E, Cozzi E. Progress in clinical encapsulated islet xenotransplantation. *Transplantation*. 2016;100(11):2301–2308.
19. Mourad NI, Perota A, Xhema D, Galli C, Gianello P. Transgenic expression of glucagon-like peptide-1 (GLP-1) and activated muscarinic receptor (M3 R) significantly improves pig islet secretory function. *Cell Transplant*. 2017;26(5):901–911.
20. Mourad NI, Gianello P. Long-term culture and in vitro maturation of macroencapsulated adult and neonatal porcine islets. *Xenotransplantation*. 2019;26(2):e12461.
21. Dufrane D, D'hoore W, Goebbels RM, Saliez A, Guiot Y, Gianello P. Parameters favouring successful adult pig islet isolations for xenotransplantation in pig-to-primate models. *Xenotransplantation*. 2006;13(3):204–214.
22. Ricordi C, Finke EH, Lacy PE. A method for the mass isolation of islets from the adult pig pancreas. *Diabetes*. 1986;35(6):649–653.
23. Gray DWR, Sudhakaran N, Titus TT, McShane P, Johnson P. Development of a novel digestion chamber for human and porcine islet isolation. *Transplant Proc*. 2004;36(4):1135–1138.
24. Li S, Sakai T, Suzuki Y, Goto T, Tanaka T, Yoshikawa T, Kakinoki K, Tanioka Y, Matsumoto I, Fujino Y, Kuroda Y. Improved quantity and in vivo function of islets isolated by reduced pressure-controlled injection of collagenase in a rat model. *Cell Transplant*. 2007;16(5):539–545.
25. Stiegler P, Stadlbauer V, Hackl F, Schaffellner S, Iberer F, Greilberger J, Strunk D, Zelzer S, Lackner C, Tscheliessnigg K. Prevention of oxidative stress in porcine islet isolation. *J Artif Organs*. 2010;13(1):38–47.